

Review Article

IRISIN: A NEWLY DISCOVERED NOVEL MYOKINE

Sana Akram¹, Hamid Javaid Qureshi²

ABSTRACT

The name irisin was given after the Greek messenger goddess Iris. It is secreted by muscles during exercise. Irisin enhances lipolysis and inhibits hepatic cholesterol synthesis. Acute exercise markedly increases irisin levels and is correlated mainly with ATP levels.

A positive correlation exists between plasma irisin and muscle mass, glucose, ghrelin, IGF1, and BMI. Plasma irisin levels are reduced markedly in patients with type 2 diabetes. Irisin levels are negatively associated with age, serum cholesterol, serum insulin, and serum adiponectin levels. Irisin induces the browning of subcutaneous white adipocytes. It increases aerobic capacity. In the liver, irisin promotes the synthesis of glycogen and inhibits gluconeogenesis. It may contribute to the beta cells profile ration. Irisin has an antioxidant effect. It induces endothelial angiogenesis.

Key Words: Exercise, BMI, Muscle

doi: <https://doi.org/10.51127/JAMDCV4I1RA01>

How to cite this:

Akram S, Qureshi HJ. Irisin: A newly discovered novel myokine. JAMDC. 2022;4(1): 31-37

doi: <https://doi.org/10.51127/JAMDCV4I1RA01>

STRUCTURE OF IRISIN

Bostrom et al 2012 for the first time identified it while searching for secretions by muscle in response to PGC-1 α activation having systemic metabolic effects.¹ PGC-1 α has been established as a major regulator of many good remodeling effects induced by exercise in muscle.²

Irisin is a 112 amino acid containing polypeptide having a molecular weight of 12 kDa. X-ray crystallography has revealed a structure that has an N-terminal domain and a C-terminal tail.³ FNIII domains are found in the extracellular domains of many receptors.⁴ Human and mouse irisin has 100% structural homology.

Fibronectin type II domain that contains protein 5 (FNDC5) in skeletal muscle secretes irisin. Expression of FNDC5 is stimulated in muscle by PGC-1 α in response to exercise.

FNDC5 is a glycosylated type I membrane protein with a 29-amino acid signal sequence at its N terminal, followed by the irisin segment, a linking segment, a transmembrane domain, and a 39 amino acid cytoplasmic domain. Cleavage in the linking peptide by an unknown protease releases irisin into the extracellular milieu.³ (Figure 1).

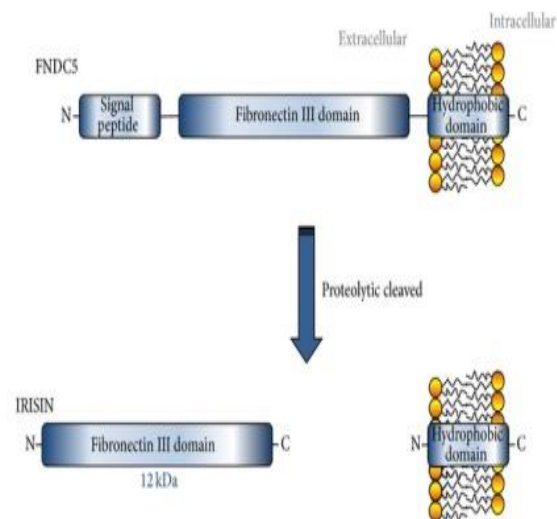


Figure 1. Structure of FNDC5 and irisin secretion.⁵

¹Assistant Professor Physiology, Faisalabad Medical College, Faisalabad.

²Professor of Physiology, Akhtar Saeed Medical & Dental College, Lahore.

MECHANISM OF ACTION

Irisin can bind to proteins of the αV class of integrins.⁶ Zhang et al. elucidated, that r-irisin administration increases UCP1 in adipose tissue through the phosphorylation of p38 mitogen-activated protein kinase (p38 MAPK) and extracellular signal-related kinase (ERK) signaling pathways.⁷ Moreover, FNDC5 overexpression promotes Akt phosphorylation, which is known to mediate the effects of insulin on glucose metabolism.⁸ Irisin has also been shown to increase lipid breakdown via the cAMP–PKA–HSL/perilipin pathway.⁹ Irisin inhibits hepatic cholesterol synthesis through successive activation of AMPK and downregulation of SREBP.¹⁰

DURATION OF ACTION

Acute exercise significantly raises irisin levels and has been correlated mainly with ATP levels. The raised irisin level during exercise has been shown to fall 30 minutes after the end of the exercise. The strong short-term effect of the irisin is to restore adenosine triphosphate (ATP) and once obtained, it decreases to basal level.¹¹

TISSUES DISTRIBUTION OF FNDC5/IRISIN GENE

Tissue-specific studies have shown that the gene for FNDC5 is predominantly expressed in human skeletal muscle. Sufficiently high expression is also present in cardiac muscle and smooth muscle of cerebral arteries and the terminal part of the colon.¹²

Low levels of FNDC5 mRNA have been demonstrated in vital organs like the kidney, liver, lung, and fat.¹² Even though sc-WAT is one of the main targets of irisin action, its expression in adipose tissue is about 1/100th the expression in muscle.

FACTORS AFFECTING IRISIN LEVELS

Plasma irisin levels are positively related to muscle mass, glucose, ghrelin, IGF-1, and BMI. A study revealed that levels of irisin were significantly declined after bariatric

surgery, indicating its positive relationship with BMI.¹¹

On the other hand, irisin levels are negatively associated with age, cholesterol, insulin, and adiponectin levels.¹¹ Irisin levels are greater in young male athletes than in middle-aged obese women.

Marked reductions in irisin levels have been reported in patients with type 2 diabetes, especially those having complications.^{13,14} Insulin resistance completely changes the metabolism of myocytes, and secretion of myokines during exercise in type 2 diabetics.¹⁵

PHYSIOLOGICAL ACTIONS OF IRISIN

Irisin induces browning of subcutaneous white adipocytes:

There is a wide difference between white adipose tissue (WAT) and brown adipose tissue (BAT) in their metabolic functions. WAT is a storehouse of triglycerides whereas BAT promotes heat production and energy consumption by expressing UCP1.⁶ Previously, BAT was thought to only help in body temperature regulation in infants and did not have much significance in adults. However, WAT contains cells that have the potential to greatly express UCP1 and transform into bright/beige cells that are adaptive brown cells. Moreover, these adaptive brown cells present in WAT are from a different lineage than the BAT. For the browning of fat, exercise is the most important factor, other factors include (1) beta-amino isobutyric acid (2) gamma-aminobutyric acid (3) PPAR γ agonists (4) JAK inhibition (5) Irisin.¹⁶

Irisin intermediate the advantageous effects of physical activity on metabolism by triggering 'Brite fat' development in subcutaneous adipocytes.¹ (Figure 2). The browning of sc-WAT is mediated by irisin-induced phosphorylation of the p38 mitogen-activated protein kinase and extracellular signal-related kinase (ERK) signaling pathways.⁷ Irisin increases energy expenditure, promotes weight loss, and

decreases insulin resistance due to fat accumulation.

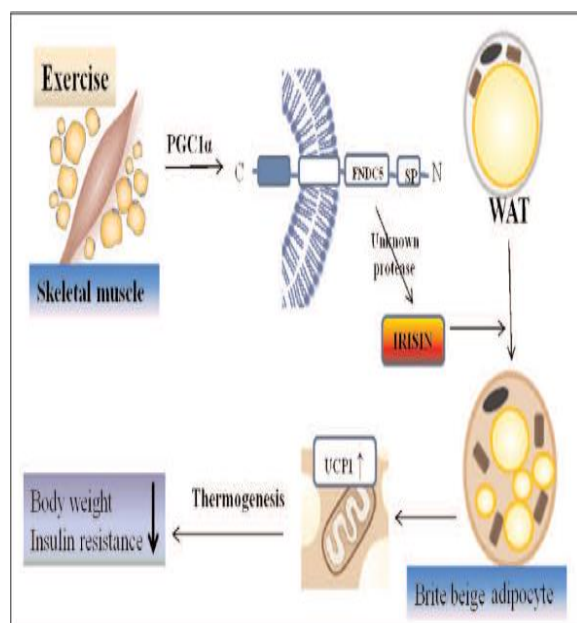


Figure 2: Irisin-induced browning of white adipocytes.¹⁶

Irisin increases aerobic capacity:

The FNDC5 and PGC-1 α genes expression has been correlated with exercise through maximal oxygen uptake (VO₂max) and gas exchange (VE/Vco₂). Between PGC-1 α and FNDC5 genes and exercise a significant positive correlation was obtained.¹⁷

Irisin promotes hepatic glycogen synthesis and inhibits gluconeogenesis:

In an STZ-high fat diabetic mouse model, continuous subcutaneous infusion of irisin improved insulin response, reduced FBG levels, increased glycogen synthase kinase 3 and Akt phosphorylation, glycogen content, and irisin level. It also inhibited phosphorylation of glycogen synthase and glucose-6-phosphatase expression in the liver.⁷ It was shown that the effects of irisin to reduce gluconeogenesis were through PI3K-Akt-FOXO1 mediated downregulation of phosphoenolpyruvate carboxykinase (PEPCK) and glucose-6 phosphatase and to increase glycogenesis via activation of glycogen synthase,⁷ thus indicating that irisin may be effective in decreasing insulin resistance in type 2 diabetes. (Figure 3)

Irisin-betatrophin axis may contribute to β cell proliferation

Irisin administration stimulates the expression of an adipokine, betatrophin, which has been shown to promote pancreatic β -cell proliferation.¹⁸ This finding suggests that increased betatrophin expression by irisin may partly explain its anti-diabetic effect. This association is, however, still controversial and needs further validation.

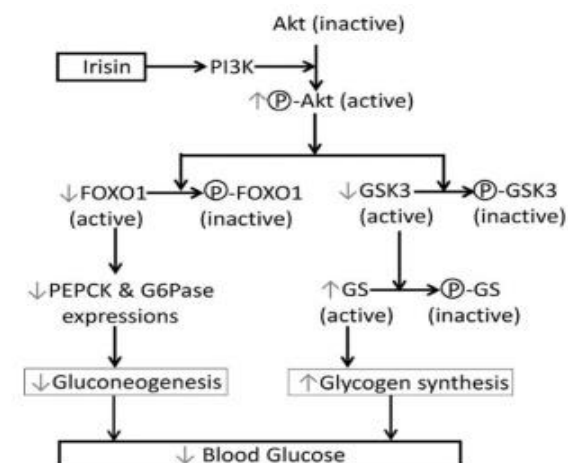


Figure 3: Effect of irisin on hepatic glucose output.⁵

Irisin's effect on lipid metabolism:

The effect of irisin on the metabolism of fat has been reported to be controversial. Most studies reported a negative correlation between irisin and lipid derangement. Increased expression of FNDC5 decreases plasma lipids and glucose in obese mice.^{9,10} In a Chinese population, a positive correlation was found between serum irisin and total cholesterol, LDL cholesterol, and free fatty acids. Reduction of irisin due to energy restriction was associated with a decline in total cholesterol and HDL cholesterol.¹⁹⁻²¹

In a study conducted by Tang et al, infusion of irisin for two weeks resulted in a reduction of plasma and hepatic cholesterol in HFD-fed obese mice. It was shown that these effects were induced by successive activation of AMPK and downregulation of SREBP2.¹³ (Figure 4). SREBP2 is a transcription factor responsible for the

activation of genes involved in cholesterol synthesis.

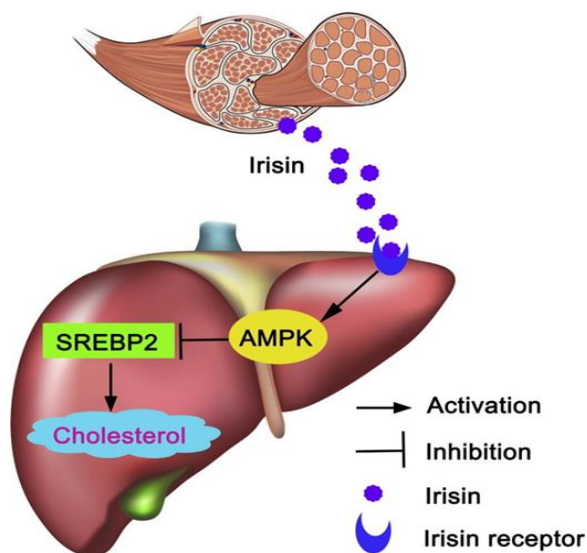


Figure 4. Effect of Irisin on hepatic cholesterol synthesis.¹⁰

Anti-oxidant effect of irisin:

Irisin, by reducing oxidative stress, has been shown to improve endothelial dysfunction and protect against atherosclerosis. Recombinant irisin significantly controlled atherosclerosis in apolipoprotein E-deficient mice.²² There was a significant reduction in ox-LDL activated biomarkers infiltration of macrophages and T cells within the atherosclerotic plaque by reducing the expression of inflammatory biomarkers. The effect of irisin on human umbilical vein endothelial cells has also revealed that irisin improves endothelial function by suppressing hyperglycemia-induced apoptosis and oxidative stress; along with increasing the expression of antioxidant enzymes.²³

Irisin induces endothelial angiogenesis:

Recent findings by Fei Wu and colleagues have shown that irisin promotes angiogenesis in a human umbilical vein through stimulation of the extracellular signal regulator kinase (ERK) pathway, proposing that irisin may have a pivotal role in preserving endothelial homeostasis.²⁴

Irisin as a biomarker of CVD in diabetes:

In patients with T2DM with macrovascular complications, irisin levels were lower as compared to patients without macrovascular complications, implying that this myokine may be a biomarker of CVD disease in people with T2DM.¹⁴

Irisin's possible role in the action of metformin:

In a study by Li and colleagues, metformin, promoted the release of irisin from skeletal muscle²⁵, suggesting that its release could be one of its mechanisms of action.

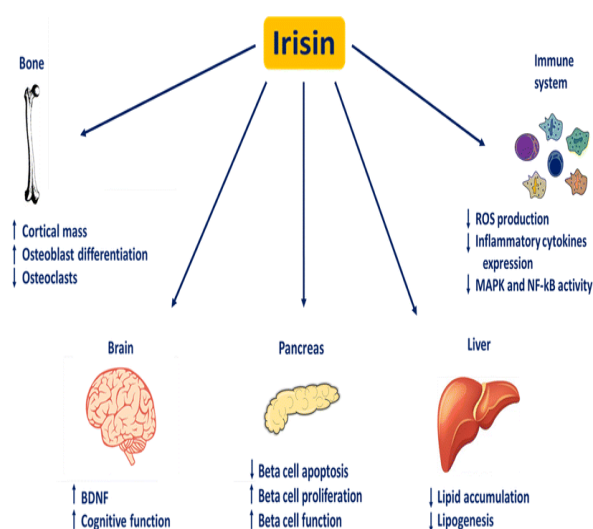
Other effects of irisin:

On bone: Increased cortical bone mineral density, thickness, and bending strength results when low dose recombinant irisin is given. (Figure 7).²⁶ This action has been attributed to activation of osteoblasts and a parallel reduction in osteoclast number and activity.

On brain: Recombinant irisin administration has been shown to promote hippocampal neurogenesis in mice, proposing its possible role in improving memory.²⁷ FNDC5 expression was shown to increase in the hippocampi of mice taking part in a thirty-day voluntary wheel running regime. FNDC5 expression in the brain upregulates the production of the neurotrophin BDNF,²⁸ which may have a role in mediating its beneficial effects in neural tissue (Figure 5).

Anti-aging effect: Plasma irisin levels showed a significant association with telomere length.²⁹ It is known that irisin signaling activates p38 MAPK which controls the expression of human telomerase reverse transcriptase.³⁰

Anti-cancer effect: According to a recent study, irisin has an inhibitory effect on the amount and metastatic properties of neoplastic breast cells³¹ and has also been shown to suppress proliferation in lung carcinoma.³¹



BDNF: brain-derived neurotrophic factor; ROS: reactive oxygen species; MAPK: mitogen-activated protein kinase; NF-κB: nuclear factor-kappa B.

Figure 5. Physiological actions of Irisin on bone, brain, pancreas, liver, and immune system.

AUTHOR'S CONTRIBUTION

SA: Manuscript writing

HJQ: Supervision and critical review

REFERENCES

- Boström P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, Rasbach KA, Boström EA, Choi JH, Long JZ, Kajimura S. A PGC1- α -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature*. 2012 Jan;481(7382):463-8. <https://doi.org/10.1038/nature10777>.
- Ferraro E, Giammarioli AM, Chiandotto S, Spoletini I, Rosano G. Exercise-induced skeletal muscle remodeling and metabolic adaptation: redox signaling and role of autophagy. *Antioxidants & redox signaling*. 2014 Jul 1;21(1):154-76.. <https://doi.org/10.1089/ars.2013.5773>.
- Schumacher MA, Chinnam N, Ohashi T, Shah RS, Erickson HP. The structure of Irisin reveals a novel intersubunit β -sheet fibronectin type III (FNIII) dimer: Implications for receptor activation. *J Biol Chem*. 2013;288:33738-44.
- Schwarzbauer JE, DeSimone DW. Fibronectins, their fibrillogenesis, and in vivo functions. *Cold Spring Harbor perspectives in biology*. 2011 Jul 1;3(7):a005041..
- Gizaw M, Anandakumar P, Debela T. A review on the role of irisin in insulin resistance and type 2 diabetes mellitus. *J. Pharmacopunct* 2017 Dec;20(4):235.. doi: 10.3831/KPI.2017.20.029
- Kim H, Wrann CD, Jedrychowski M, Vidoni S, Kitase Y, Nagano K, Zhou C, Chou J, Parkman VJ, Novick SJ, Strutzenberg TS. Irisin mediates effects on bone and fat via α V integrin receptors. *Cell*. 2018 Dec 13;175(7):1756-68. <https://doi.org/10.1016/j.cell.2018.10.025>
- Zhang Y, Li R, Meng Y, Li S, Donelan W, Zhao Y, Qi L, Zhang M, Wang X, Cui T, Yang LJ. Irisin stimulates browning of white adipocytes through mitogen-activated protein kinase p38 MAP kinase and ERK MAP kinase signaling. *Diabetes*. 2014 Feb 1;63(2):514-25. <https://doi.org/10.2337/db13-1106>.
- Liu TY, Shi CX, Gao R, Sun HJ, Xiong XQ, Ding L, Chen Q, Li YH, Wang JJ, Kang YM, Zhu GQ. Irisin inhibits hepatic gluconeogenesis and increases glycogen synthesis via the PI3K/Akt pathway in type 2 diabetic mice and hepatocytes. *Clin sci*. 2015 Nov 1;129(10):839-50. <https://doi.org/10.1042/CS20150009>.
- Xiong XQ, Chen D, Sun HJ, Ding L, Wang JJ, Chen Q, Li YH, Zhou YB, Han Y, Zhang F, Gao XY. FNDC5 overexpression and irisin ameliorate glucose/lipid metabolic derangements and enhance lipolysis in obesity. *Biochim Biophys Acta Mol Basis Dis*. 2015 Sep 1;1852(9):1867-75. <https://doi.org/10.1016/j.bbdis.2015.06.017>.
- Tang H, Yu R, Liu S, Huwatibieke B, Li Z, Zhang W. Irisin inhibits hepatic cholesterol synthesis via AMPK-SREBP2 signaling. *EBioMedicine*. 2016 Apr 1;6:139-48..
- Huh JY, Panagiotou G, Mougios V, Brinkoetter M, Vamvini MT, Schneider BE, Mantzoros CS. FNDC5 and irisin in humans: I. Predictors of circulating concentrations in serum and plasma and II. mRNA expression and circulating concentrations in response to weight loss and exercise. *Metab*. 2012 Dec 1;61(12):1725-38. <https://doi.org/10.1016/j.metabol.2012.09.002>.
- Martinez Munoz IY, Camarillo Romero ED, Garduno Garcia JD. Irisin a novel metabolic biomarker: present knowledge and future directions *IJED*. 2018 Oct 9;2018. <https://doi.org/10.1155/2018/7816806>.

13. Liu JJ, Wong MD, Toy WC, Tan CS, Liu S, Ng XW, Tavintharan S, Sum CF, Lim SC. Lower circulating irisin is associated with type 2 diabetes mellitus. *JDC* 2013 Jul 1;27(4):365-9. <https://doi.org/10.1016/j.jdiacomp.2013.03.002>.
14. Zhang M, Chen P, Chen S, Sun Q, Zeng QC, Chen JY, Liu YX, Cao XH, Ren M, Wang JK. The association of new inflammatory markers with type 2 diabetes mellitus and macrovascular complications: a preliminary study. *Eur Rev Med Pharmacol Sci*. 2014 Jun 1;18(11):1567-72..
15. Smith U, Kahn BB. Adipose tissue regulates insulin sensitivity: role of adipogenesis, de novo lipogenesis and novel lipids. *J. Intern. Med.*. 2016 Nov;280(5):465-75..
16. Lecker SH, Zavin A, Cao P, Arena R, Allsup K, Daniels KM, Joseph J, Schulze PC, Forman DE. Expression of the irisin precursor FNDC5 in skeletal muscle correlates with aerobic exercise performance in patients with heart failure. *Circ Heart Fail*. 2012;5:812-8 <https://doi.org/10.1111/joim.12540>
17. Sanchis-Gomar F, Perez-Quilis C. The p38–PGC-1 α –irisin–betatrophin axis: Exploring new pathways in insulin resistance. *Adipocyte*. 2014 Jan 28;3(1):67-8.. <https://doi.org/10.4161/adip.27370>
18. Park MJ, Kim DI, Choi JH, Heo YR, Park SH. New role of irisin in hepatocytes: The protective effect of hepatic steatosis in vitro. *Cellular signalling*. 2015 Sep 1;27(9):1831-9. <https://doi.org/10.1016/j.cellsig.2015.04.010>
19. Tang S, Zhang R, Jiang F, Wang J, Chen M, Peng D, Yan J, Wang S, Bao Y, Hu C, Jia W. Circulating irisin levels are associated with lipid and uric acid metabolism in a Chinese population. *Clin. Exp. Pharmacol.*. 2015 Sep;42(9):896-901. <https://doi.org/10.1111/1440-1681.12439>.
20. Chang CL, Huang SY, Soong YK, Cheng PJ, Wang CJ, Liang IT. Circulating irisin and glucose-dependent insulinotropic peptide are associated with the development of polycystic ovary syndrome. *J Clin Endocrinol Metab* 2014 Dec 1;99(12):E2539-48. <https://doi.org/10.1210/jc.2014-1180>.
21. Lu J, Xiang G, Liu M, Mei W, Xiang L, Dong J. Irisin protects against endothelial injury and ameliorates atherosclerosis in apolipoprotein E-Null diabetic mice. *Atherosclerosis*. 2015 Dec 1;243(2):438-48. <https://doi.org/10.1016/j.atherosclerosis.2015.10.020>
22. Zhu DI, Wang H, Zhang J, Zhang X, Xin C, Zhang F, Lee Y, Zhang L, Lian K, Yan W, Ma X. Irisin improves endothelial function in type 2 diabetes through reducing oxidative/nitrative stresses. *J Mol Cell Cardiol*. 2015 Oct 1;87:138-47. <https://doi.org/10.1016/j.yjmcc.2015.07.015>.
23. Wu F, Song H, Zhang Y, Zhang Y, Mu Q, Jiang M, Wang F, Zhang W, Li L, Li H, Wang Y. Irisin induces angiogenesis in human umbilical vein endothelial cells in vitro and in zebrafish embryos in vivo via activation of the ERK signaling pathway. *PLoS one*. 2015 Aug 4;10(8):e0134662.. <https://doi.org/10.1371/journal.pone.0134662>
24. Li DJ, Huang F, Lu WJ, Jiang GJ, Deng YP, Shen FM. Metformin promotes irisin release from murine skeletal muscle independently of AMP-activated protein kinase activation. *Acta physiologica*. 2015 Mar;213(3):711-21. <https://doi.org/10.1111/apha.12421>.
25. Colaianni G, Cuscito C, Mongelli T, Pignataro P, Buccoliero C, Liu P, Lu P, Sartini L, Di Comite M, Mori G, Di Benedetto A. The myokine irisin increases cortical bone mass. *Proceedings of the National Academy of Sciences*. 2015 Sep 29;112(39):12157-62. <https://doi.org/10.1073/pnas.151662211>.
26. Wrann CD, White JP, Salogiannis J, Laznik-Bogoslavski D, Wu J, Ma D, Lin JD, Greenberg ME, Spiegelman BM. Exercise induces hippocampal BDNF through a PGC-1 α /FNDC5 pathway. *Cell Metab.*. 2013 Nov 5;18(5):649-59. <https://doi.org/10.1016/j.cmet.2013.09.008>
27. Islam MR, Young MF, Wrann CD. The Role of FNDC5/Irisin in the Nervous System and as a Mediator for Beneficial Effects of Exercise on the Brain. *Hormones, metabolism and the benefits of exercise*. 2017:93-102. https://doi.org/10.1007/978-3-319-72790-5_8
28. Rana KS, Arif M, Hill EJ, Aldred S, Nagel DA, Nevill A, Randeva HS, Bailey CJ, Bellary S, Brown JE. Plasma irisin levels predict telomere length in healthy adults. *Age*. 2014 Apr;36(2):995-1001 <https://doi.org/10.1007/s11357-014-9620-9>.

29. Matsuo T, Shimose S, Kubo T, Fujimori J, Yasunaga Y, Sugita T, Ochi M. Correlation between p38 mitogen-activated protein kinase and human telomerase reverse transcriptase in sarcomas. *J. Exp. Clin. Cancer Res.* 2012 Dec;31(1):1-9. <https://doi.org/10.1186/1756-9966-31-5>.
30. Gannon NP, Vaughan RA, Garcia-Smith R, Bisoffi M, Trujillo KA. Effects of the exercise-inducible myokine irisin on malignant and non-malignant breast epithelial cell behavior in vitro. *Int J Cancer.* 2015 Feb 15;136(4):E197-202. <https://doi.org/10.1002/ijc.29142>.
31. Shao L, Li H, Chen J, Song H, Zhang Y, Wu F, Wang W, Zhang W, Wang F, Li H, Tang D. Irisin suppresses the migration, proliferation, and invasion of lung cancer cells via inhibition of epithelial-to-mesenchymal transition. *Biochem Biophys Res Commun* 2017 Apr 8;485(3):598-605. <https://doi.org/10.1016/j.bbrc.2016.12.084>.